

Note

Electron-impact and chemical-ionization mass-spectral identification of methylated derivatives obtained from 2-acetamido-2,4-dideoxy- and 3-acetamido-3,4-dideoxy-DL-pentopyranoses

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Mass spectrometry has become one of the most powerful techniques for structural investigations of complex carbohydrates^{1,2}.

Interest in the synthesis and the structural characterization of novel amino sugars has provided a stimulus to maintain the need for systematic mass-spectral studies of series of amino sugars.

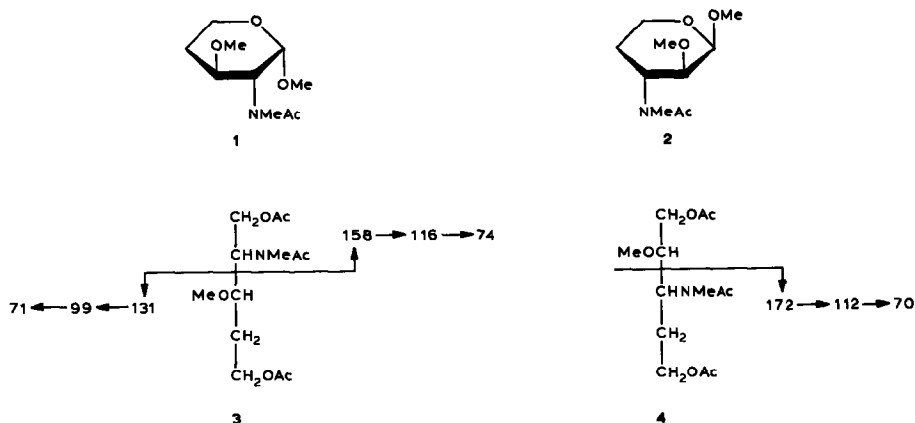
The unusual and novel amino sugars 2-amino-2,4-dideoxy-DL-pentose and 3-amino-3,4-dideoxy-DL-pentose constitute the glycosyl portion of various 2'- and 3'-aminodideoxyribonucleosides that have been shown to exhibit antiviral and anti-tumor properties^{3,4}.

In a continuation of our studies on the mass spectrometry of some acetylated derivatives of these novel aminodideoxypentoses⁵, we now report the electron-impact and chemical-ionization mass-spectra of some permethylated glycosides of, and methylated alditol acetates from, 2-acetamido-2,4-dideoxy- and 3-acetamido-3,4-dideoxy-DL-pentoses.

The electron-impact mass spectra of methyl 2,4-dideoxy-3-O-methyl-2-(N-methylacetamido)- β -DL-*threo*-pentopyranoside (1) and methyl 3,4-dideoxy-2-O-methyl-3-(N-methylacetamido)- β -DL-*threo*-pentopyranoside (2) are shown in Table I, and the proposed modes of formation of the specific ions formed during the break-

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down of the molecular radical-ion are identical to the ones obtained for the fully acetylated derivatives of 1 and 2 (see Ref. 5). The e.i.-mass spectra of *O*-(trideuteriomethyl)ated and *N*-(trideuterioacetyl)ated analogs of 1 and 2 support this conclusion.

The isobutane chemical-ionization mass spectrum of the methyl glycoside 1 gave, *inter alia*, peaks at the following *m/z* values: 144 (5.1), 154 (12.5), 186 (18.3) 218 (100), 219 (11.5), and 256 (2.8). The ion at *m/z* 256 is assigned to $[M + C_3H_3]^+$.

TABLE I

ELECTRON-IMPACT MASS SPECTRA AND RETENTION TIMES OF THE VARIOUS DERIVATIVES 1-4 OF 2-AMINO-2,4-DIDEOXY- AND 3-AMINO-3,4-DIDEOXY-DL-PENTOSE

Compound	<i>m/z</i> (intensity, %)	<i>T_R</i> ^a
Methyl 2,4-dideoxy-3- <i>O</i> -methyl-2-(<i>N</i> -methylacetamido)-β-DL- <i>threo</i> -pentopyranoside (1)	217(3.1), 186(3.0), 185(2.2), 170(3.3), 154(18.5), 144(41.4), 129(28.0), 126(30.2), 125(38.1), 114(39.1), 112(12.1), 99(59.5), 98(39.2), 87(100.0), 84(29.9), 75(45.1), 72(63.1), 57(49.6), 43(60.3)	0.41
Methyl 3,4-dideoxy-2- <i>O</i> -methyl-3-(<i>N</i> -methylacetamido)-β-DL- <i>threo</i> -pentopyranoside (2)	218(4.8), 202(9.7), 187(3.2), 186(10.5), 170(5.1), 157(68.4), 144(13.6), 142(100.0), 129(29.7), 114(15.0), 113(34.4), 112(28.9), 100(10.1), 99(11.1), 88(71.2), 87(51.4), 86(36.4), 84(62.4), 43(57.1)	0.43
1,5-Di- <i>O</i> -acetyl-2,4-dideoxy-3- <i>O</i> -methyl-2-(<i>N</i> -methylacetamido)-DL- <i>threo</i> -pentitol (3)	158(20.0), 149(6.4), 142(2.8), 131(21.1), 116(100.0), 99(28.8), 74(38.5), 71(49.3), 57(18.5), 43(62.1)	0.62
1,5-Di- <i>O</i> -acetyl-3,4-dideoxy-2- <i>O</i> -methyl-3-(<i>N</i> -methylacetamido)-DL- <i>threo</i> -pentitol (4)	216(3.1), 172(21.2), 112(100.0), 70(69.7), 43(23.2)	0.63

^aRetention time relative to that of D-glucitol hexaacetate as unity.

The primary fragment-ion at m/z 186 is generated by the loss of a molecule of methanol from the protonated molecular ion $[M + H]^+$ at m/z 218 (base peak). The loss of a molecule of ketene from the ion at m/z 186 produces the ion at m/z 144.

The isobutane chemical-ionization mass spectrum of the methyl glycoside 2 gave, *inter alia*, peaks of the following m/z values: 256 (2.7), 219 (10.9), 218 (100), 217 (4.2), 187 (5.0), 186 (48.5), and 185 (4.0). The ions at m/z 256 and m/z 186 are assigned as before. It should be noted that the ion at m/z 186 does not lose a molecule of ketene, contrary to its corresponding ion in the chemical-ionization mass spectrum of the isomeric methyl glycoside 1.

The simple and well established behavior of partially methylated alditol acetates upon electron impact^{2,6} makes these derivatives suitable for the identification of these novel aminodideoxypentoses.

The electron-impact mass spectra of 1,5-di-*O*-acetyl-2,4-dideoxy-3-*O*-methyl-2-(*N*-methylacetamido)-*DL*-*threo*-pentitol (3) and 1,5-di-*O*-acetyl-3,4-dideoxy-2-*O*-methyl-3-(*N*-methylacetamido)-*DL*-*threo*-pentitol (4), given in Table I, were found to obey the same fragmentation pattern as other partially methylated alditol acetates of amino sugars⁶. As expected, the fragmentation pattern of alditol derivatives 3 and 4 was governed by fission between C-2 and C-3 of the alditol chains, to afford the primary fragment-ions at m/z 158 and 172, respectively. The breakdown processes leading to the production of the aforementioned fragment-ions were investigated by *N*-(trideuterioacetyl)ation. In effect, the corresponding *N*-[methyl(trideuterioacetyl)]-ated derivatives of 3 and 4 gave a fragmentation pattern analogous to that of their precursors, and consequently, the ions at m/z 158 and 172 shifted to three a.m.u. higher respectively.

From Table I, it may be seen that the relative retention time cannot be depended on as a diagnostic tool for the identification of this series of reported derivatives. This makes it essential that the final identification be made by electron-impact mass spectrometry.

In conclusion, the relevant data obtained from the electron-impact and chemical-ionization mass spectra provide valuable information for the clear identification of this novel series of reported derivatives of aminodideoxypentoses.

EXPERIMENTAL

Reagents. — All of the reagents and solvents were of analytical grade, were glass-distilled before use, and were stored over molecular sieves 4A.

Synthesis of the methyl glycosides. — The methyl 2,4-dideoxy-3-*O*-methyl-2-(*N*-methylacetamido)- β -*DL*-*threo*-pentopyranoside (1) and methyl 3,4-dideoxy-2-*O*-methyl-3-(*N*-methylacetamido)- β -*DL*-*threo*-pentopyranoside (2) were obtained from the fully acetylated precursor derivatives⁵ by methylation of the corresponding acetylated methyl glycosides by the Hakomori method⁷ and were purified by passage through a column of Sephadex LH-20.

Synthesis of the alditol acetates. — The methyl glycosides 1 and 2 were hydro-

lyzed with 0.5M trifluoroacetic acid for 1 h at 100°, and the solutions were evaporated to dryness. The free dideoxyamino sugars resulting were reduced with sodium borohydride in water for 1 h at room temperature, the base neutralized with dilute acetic acid, the solution evaporated to dryness, and traces of solvents co-distilled with methanol-acetic acid. Each alditol was acetylated with 1:1 acetic anhydride-pyridine for 1 h at 100°, and the solution evaporated to afford, respectively, 1,5-di-*O*-acetyl-2,4-dideoxy-3-*O*-methyl-2-(*N*-methylacetamido)-DL-*threo*-pentitol (3) and 1,5-di-*O*-acetyl-3,4-dideoxy-2, *O*-methyl-3-(*N*-methylacetamido)-DL-*threo*-pentitol (4).

Gas-liquid chromatography. — Gas-liquid chromatography was performed on a fused-silica capillary column (0.25 × 0.23 mm, i.d., film thickness 0.15 μm) of WCOT CP-Sil 5CB (Chrompack), at 150°, mounted in a Perkin-Elmer Model 8310 gas chromatograph equipped with a flame-ionization detector.

Gas-liquid chromatography-mass spectrometry. — Combined gas-liquid chromatography-electron-impact mass spectrometry was performed in a Hewlett Packard Model 5985 A GC/MS/DS instrument equipped with a dual e.i./c.i. source. E.i. spectra were recorded at a source temperature of 160° and an ionizing voltage of 70 eV. C.i. spectra were recorded at a source pressure of 120 Pa (using isobutane as the reagent gas and carrier), a source temperature of 150°, and an ionization voltage of 230 eV. The temperature program for the e.i. and c.i. spectra started at 120° and was increased to 270° at 10°/min, using a packed glass column of 2% of OV-17 on Chromosorb W (H.P.) (80-100 mesh).

REFERENCES

1. T. RADFORD AND D. C. DEJONGH, IN G. R. WALLER (Ed.), *Biochemical Applications of Mass Spectrometry*, Wiley, New York, 1972 pp. 313-350.
2. T. RADFORD AND D. C. DEJONGH, IN G. R. WALLER (Ed.), *Biochemical Applications of Mass Spectrometry*, Wiley, New York, 1980, pp. 255-310.
3. E. DE CLERQ, J. BALZARINI, J. DESCAMPS, AND F. ECKSTEIN, *Biochem. Pharmacol.*, 29 (1980) 1849-1851.
4. D. BOUCHU, M. ABOU-ASSALI, A. GROUILLER, G. GARRETT, AND H. PACHECO, *Eur. J. Med. Chem.*, 16 (1981) 43-47.
5. J. H. BANOUB, F. MICHON, R. ROY, A. GROUILLER, AND H. BAZIN, *Carbohydr. Res.*, 144 (1985) 127-136.
6. J. LÖNNGREN AND S. SVENSSON, *Adv. Carbohydr. Chem. Biochem.*, 29 (1974) 41-106.
7. S. HAKOMORI, *J. Biochem. (Tokyo)*, 55 (1964) 205-208.